

# AN ANIMAL-FREE HYDROGEL FOR TRANSLATIONAL STUDIES *IN VITRO* – *IN VIVO* – *IN HUMAN*

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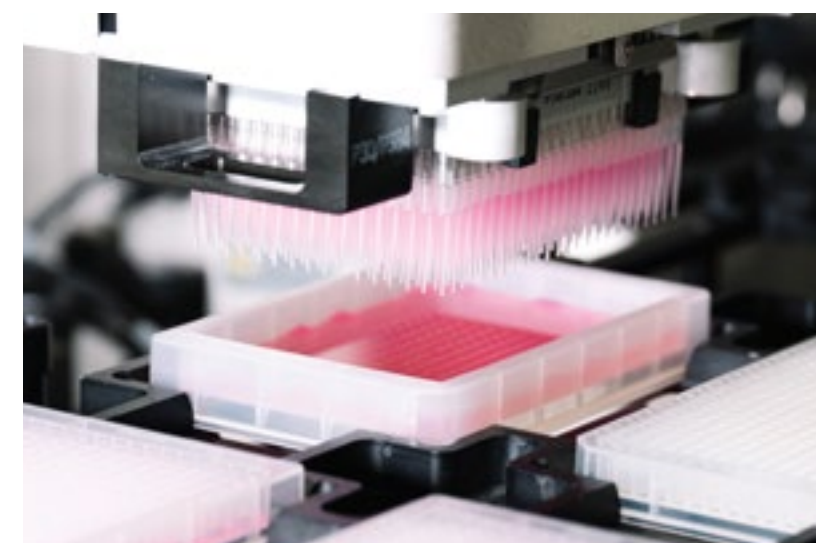
## INTRODUCTION TO NANOFIBRILLAR CELLULOSE (NFC) HYDROGEL



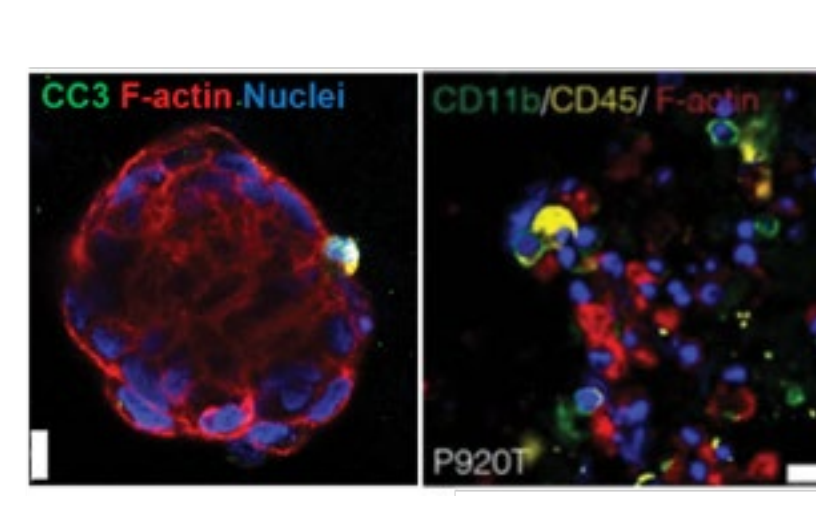
The hydrogel is **shear-thinning** and therefore can be readily dispensed, printed, injected without the need for cross-linking. The viscosity can be simply tuned by dilution.



The material is highly **bio-compatible** (200 protocols including primary cells, also tested in ISO10993). Here, SEM image of fibroblasts (\*) adhering (arrows) to the nanofibers (#). J. Sheard

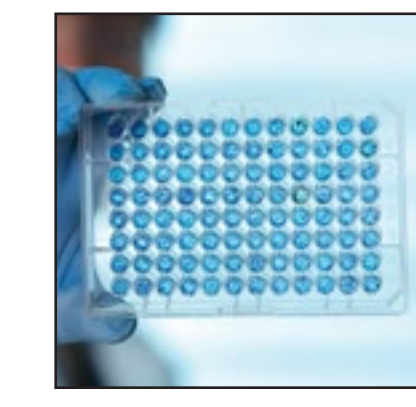


GrowDex<sup>®</sup> is room **temperature stable**, allowing automation of assays. GrowDex is proven to work with many dispensing and pipetting robots.



The biomaterial is inert and preserves the immune response of primary cells. The image shows that immune cells are retained in patient derived breast cancer explant cultures. Ref. P. Munne

## REPLACING THE INVISIBLE ANIMALS WITH WOOD



96 well plate



1 mouse



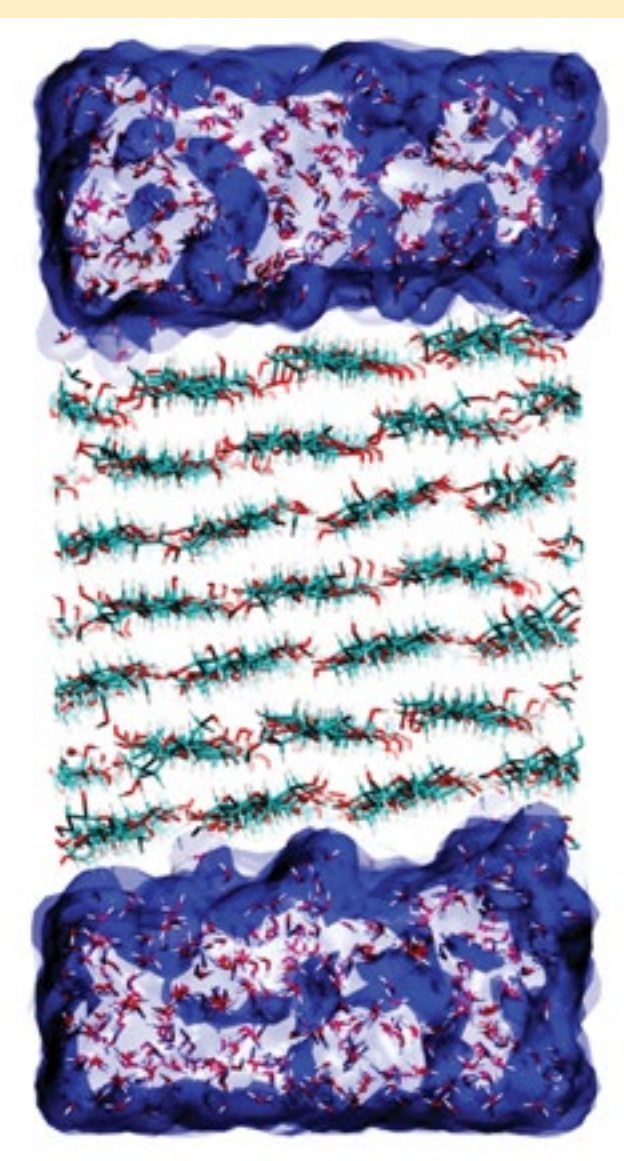
0.33 cm<sup>3</sup> wood

While implementing 3R's, many focus on the visible animals – needed for *in vivo* tests. However, many animals are used *in vitro*, e.g. in FBS, and many assays rely on a matrix isolated from mice [6]. On average, for a 96 well plate 5 ml of animal-derived matrix stock [6] or 3.3 ml of wood-derived matrix stock are needed.

UPM birch-based nanofibrillar cellulose (NFC) only consists of wood and purified water. The GrowDex<sup>®</sup> hydrogels do not contain any animal or human-derived material. The production follows ISO13485 standard.

### *In silico*

An *in silico* model of nanofibrillar cellulose has been built to enable fast testing of the hydrogel and its behaviour in the presence of different molecules and in various environments.



In the study by Koivunotko et al., the quality of the *in silico* model has been shown in the correlation of simulations and experimental data for freeze-dried and native hydrogel. [7]

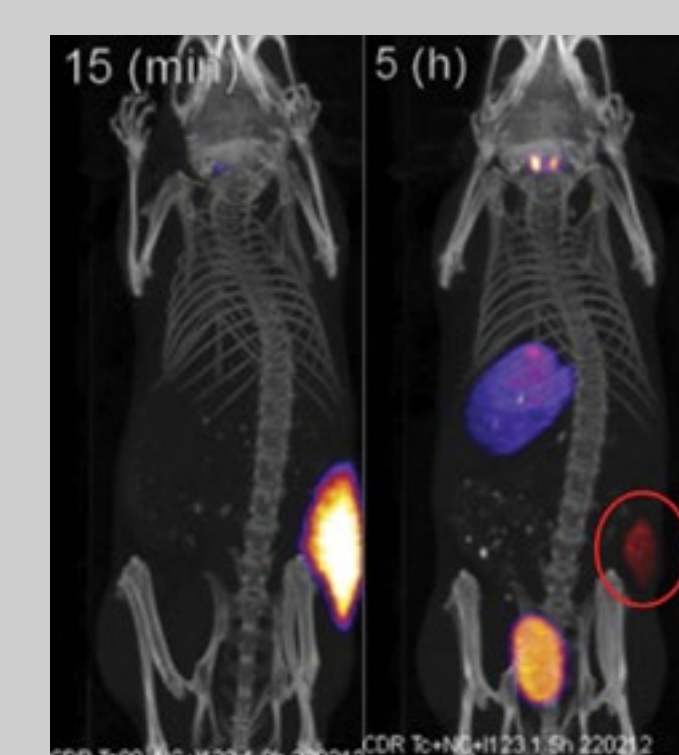
### *In vitro*

GrowDex, GrowDex-T and GrowDex-A are NFC hydrogels for 3D cell culture. GrowDase<sup>™</sup> enables the gentle release of cells for analysis. Protocols for 200 cell types have been established including cell lines as well as primary samples, e.g. a respiratory cell culture which allows live cell imaging and was maintained for up to 700 days. [2,8].

PHH and HepaRG cells retain their viability and their functionality, as shown by albumin expression and CYP activity, up to 5 weeks in GrowDex [9-11]. In addition, PHH can be co-cultured with Kupffer cells in GrowDex to mimic *in vivo* tissue morphology and functionality [10]. The applicability in automated assays and the reproducibility make GrowDex the perfect matrix for toxicology studies.

### *In vivo*

UPM's NFC is suitable as an injectable matrix for drug delivery and cell implantation. It allows controlled release and local delivery of therapeutic compounds as shown with radiolabeled model substances [3]. GrowDex-T has been used as an artificial extracellular matrix for the delivery of spheroids of otic neuronal progenitor cells into the mouse inner ear, leading to 90 days survival of the spheroids [12]. In addition, it has been shown *in vivo* that UPM's NFC does not cause any acute systemic toxicity (unpublished results).

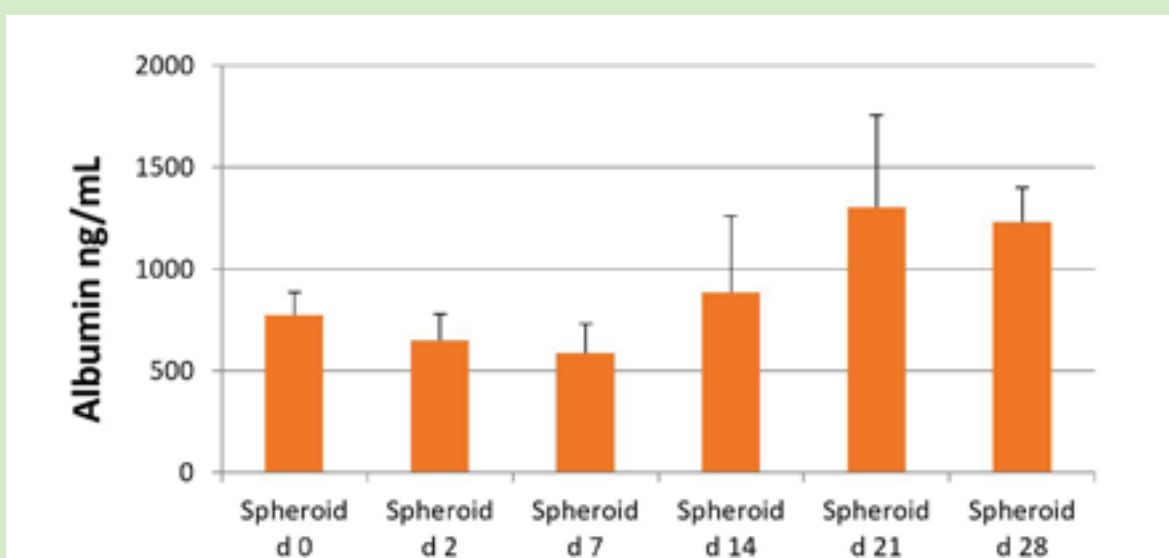


123I-Nal was found at the injection site. [3]

Dual tracing image showing the NFC implant (red) and test compound release. 123I-Nal was mostly distributed into the thyroid glands and stomach, and was excreted to urine. 5 h post injection, no trace of

### Clinical

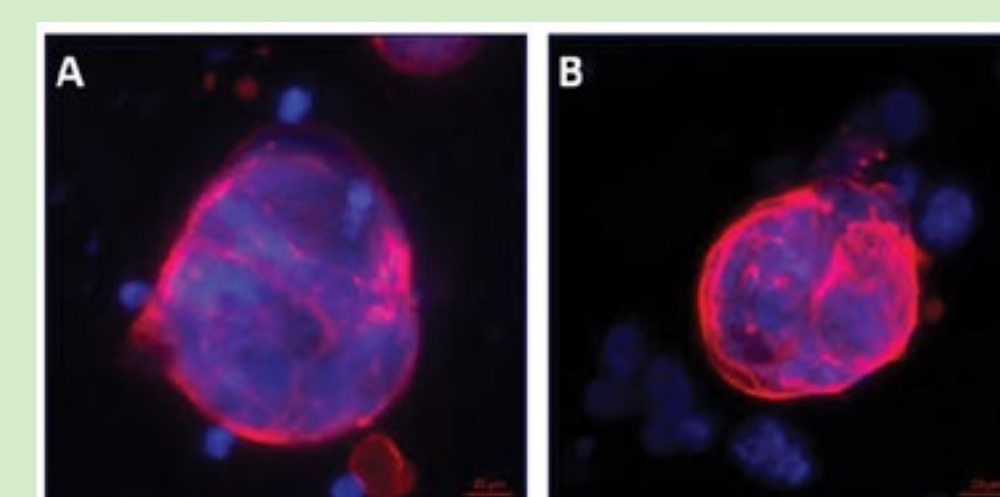
UPM's NFC is the basis for the CE-marked wound dressing FibDex<sup>®</sup>. It is intended for use on skin graft donor site wounds and offers excellent performance and healing results. As a one-time dressing, FibDex also reduces the workload for healthcare professionals as well as the waste amount in hospitals. Clinical investigation of FibDex for superficial dermal burns is on the way [13]. Moreover, the promising *in vivo* data suggest a fruitful future development of a wider spectrum of clinical applications for UPM's NFC.



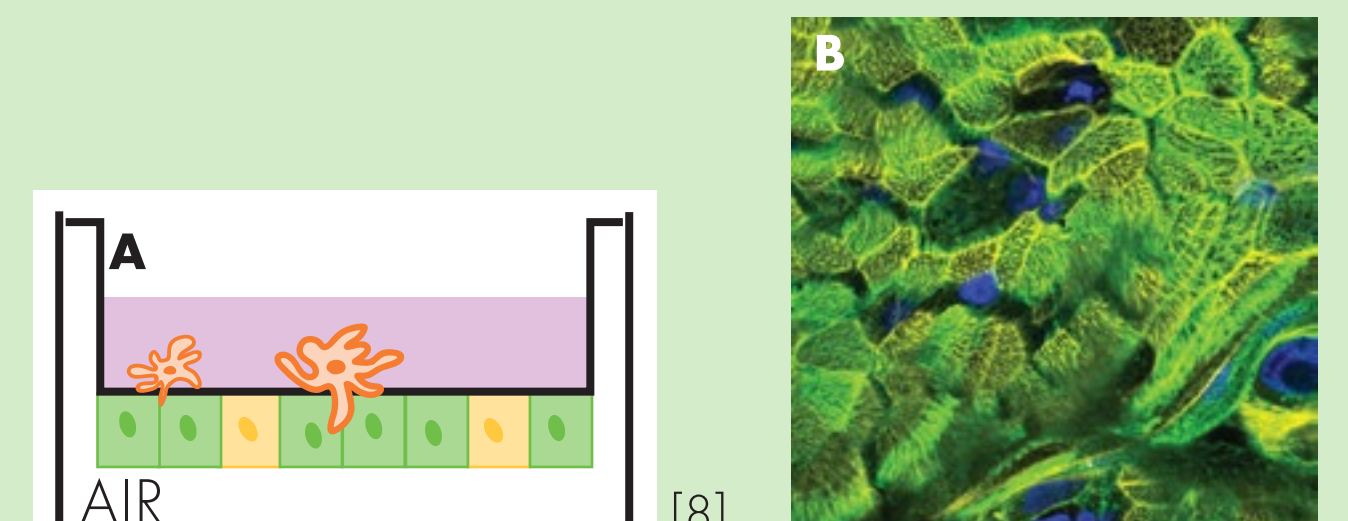
**Albumin secretion.** Primary hepatocytes were cultured for 28 days as spheroid culture (after 7d pre-culture) on GrowDex. Albumin secretion was quantified via ELISA.



**CYP3A4 activity.** Primary hepatocytes were cultured for 28 days as spheroid culture (after 7d pre-culture) on GrowDex. CYP3A4 activity was quantified by analyzing the testosterone-6 $\beta$ -hydroxylation via LC-MS [1].

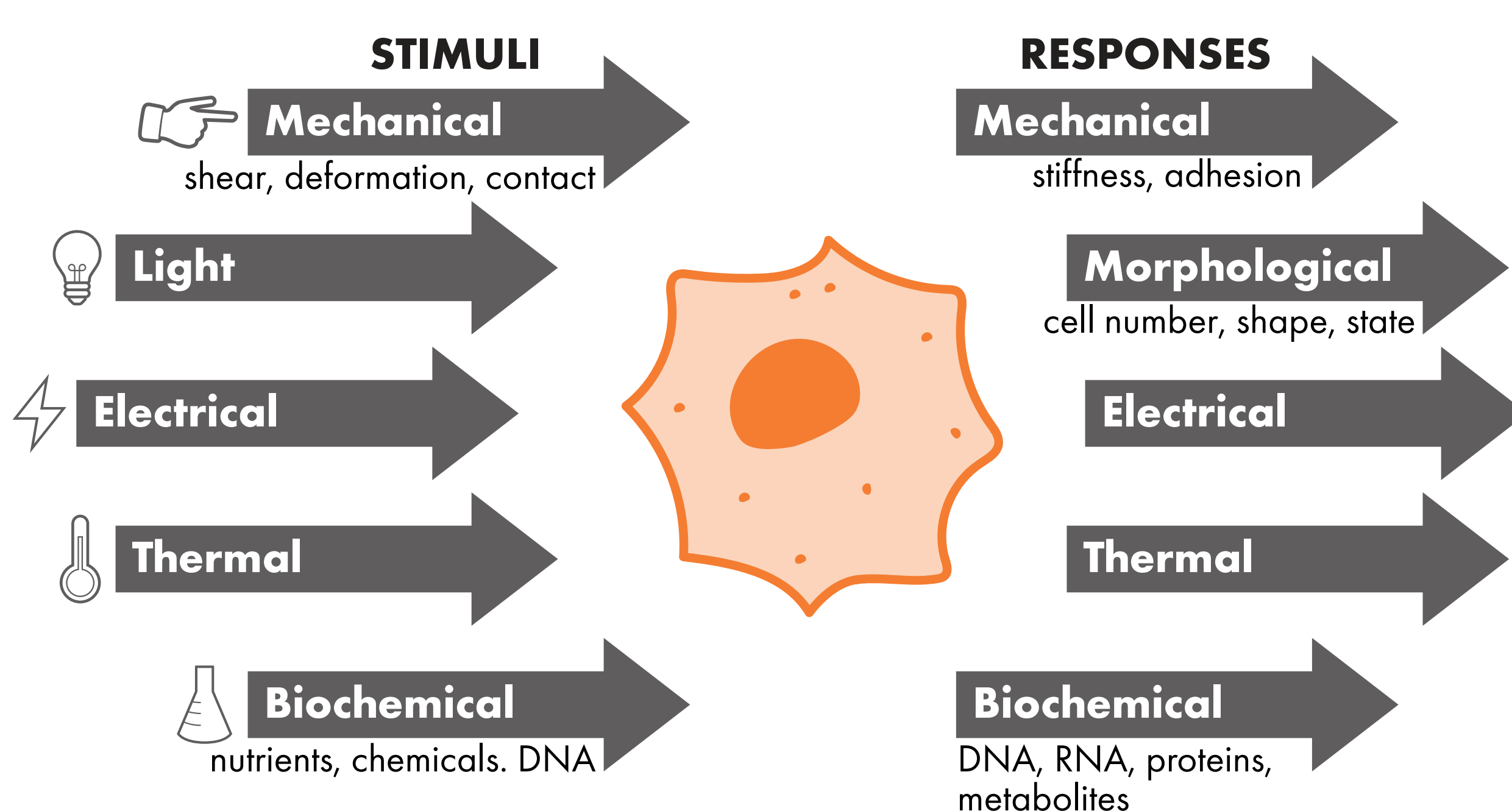


**Primary Human Hepatocyte/Kupffer cell coculture.** Spheroids stained with DAPI (blue) and Phalloidin (red) after 12 days culture (ratio PHH:Kupffer cells 2:1) in A) 0.8 % GrowDex, and B) 0.5 % GrowDex.



**Long term 3D respiratory cell culture.** A) Schematic of upside-down culture of respiratory epithelial cells seeded in GrowDex (adapted from ref. 8). This technique allows transfer of the cultures for live cell imaging and returning them to the culture well. B) Human bronchial epithelial cells were cultured in GrowDex for 700 d and stained with WGA-488 (green), Phalloidin (yellow), and Hoechst (blue) [8].

## CELLS ARE NOT REPEATABLE



In order to capture the complex cell responses we need to make everything else as repeatable as possible...

## HOW TO IMPROVE REPEATABILITY AND RELEVANCE OF CELL MODELS?

Cells	Cancer origin 2D culture	Patient-derived 3D culture
Cell delivery	Frozen Thawing	Fresh Ready-to-use
Handling	Human error	Reproducible Automated GrowDex <sup>®</sup>
Matrix	Varying composition	Reproducible No animal DNA
Results	Varying	Repeatable Reliable

## CONCLUSIONS

- Birch nanocellulose (GrowDex) is a novel inert animal-free hydrogel
- It has been shown to work across in silico – *in vitro* – *in vivo* – clinical use.
- The GrowDex hydrogel helps implementing the 3R policy:
  - Reducing use of invisible animals in *in vitro* assays (by replacing animal-derived matrix with wood)
  - Reducing use of visible animals in *in vivo* models (by replacing animal tests with cell-based assays)
  - Refining use of animals (by enabling improved models)
- Work with us:**
  - In vitro*: We are looking for partners to develop 3D *in vitro* toxicology assays.
  - In vivo*: We are looking for partners interested in cell implantation.

## REFERENCES:

- [1] Paasonen L. et al., Analysis of CYP3A4 enzyme activity for primary hepatocytes cultured in 3D, Application Note 9, UPM-Kymmene Oyj.[2] Posch W. et al., C5aR inhibition of non-immune cells suppresses inflammation and maintains epithelial integrity in SARS-CoV-2-infected primary human airway epithelia. J Allergy Clin Immunol. 2021 Jun;147(6):2083-2097.[3] Laurén P. et al., Technetium-99m-labeled nanofibrillar cellulose hydrogel for in vivo drug release. Eur J Pharm Sci. 2014 Dec 18;65:79-88.[4] Heuer R. A. et al., Three-Dimensional Otic Neuronal Progenitor Spheroids Derived from Human Embryonic Stem Cells. Tissue Eng Part A. 2021 Feb;27(3-4):256-269.[5] Koivuniemi R. et al., Clinical Study of Nanofibrillar Cellulose Hydrogel Dressing for Skin Graft Donor Site Treatment. Adv Wound Care 2020 Apr 1;9(4):199-210.[6] Kibbey M.C., Maintenance of the EHS sarcoma and Matrigel preparation. J Tissue Cult Methods 16: 227-230, 1994.[7] Koivunotko E. et al., Molecular Insights on Successful Reconstitution of Freeze-Dried Nanofibrillated Cellulose Hydrogel. ACS Applied Bio Materials 2021 Sep 20;4(9):7157-7167.[8] Zaderer V. et al., Turning the World Upside-Down in Cellulose for Improved Culturing and Imaging of Respiratory Challenges within a Human 3D Model. Cells 2019 Oct 21;8(10):1292.[9] Paasonen L. et al., Long-term 3D Culture of Human Primary Hepatocytes in GrowDex<sup>®</sup>. Application Note 7, UPM-Kymmene Oyj.[10] Paasonen L. et al., Co-Culturing of Primary Hepatocytes and Kupffer Cells in GrowDex<sup>®</sup> and DAPI/Phalloidin Staining of the Cell Spheroids. Application Note 8, UPM-Kymmene Oyj.[11] Paasonen L. et al., Analysis of CYP3A4 enzyme activity for primary hepatocytes cultured in 3D. Application Note 9, UPM-Kymmene Oyj.[12] Chang H.-T. et al., An engineered three-dimensional stem cell niche in the inner ear by applying a nanofibrillar cellulose hydrogel with a sustained-release neurotrophic factor delivery system. Acta Biomater 2020 May;108:111-127.[13] https://www.clinicaltrials.gov/study/NCT05629091.